Deep Learning Tools in NMR

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Introduction

AI, machine learning, and deep learning are interchangeable and easily confusing



Over several decades ...

- Struggle to match human performance intuitive human judgment
- Only on 'perfect datasets' sharp well-resolved with minimal noise

Deep Learning

- Automate or improve different analysis stages
- Increasing efficiency, utility and ease of use of NMR spectroscopy

Reconstruction of non-uniformly sampled (NUS)



Several excellent algorithms for reconstructing NUS NMR spectra using non-DL methods:

- SMILE (Ying et al. 2017)
- hmsIST (Hyberts et al. 2012)
- -MDD-NMR (Jaravine et al. 2006).

Recent proof-of-principles studies have shown that DL based reconstruction methods have the ability to give reconstructions more rapidly and with higher fidelity than existing methods

FID-Net A versatile DNN architecture

FID-Net works effectively:

- arbitrary sampling schedules (meaning can be deployed w/o further training & minimal user input)

- processing time domain data beyond reconstruction tasks

- Virtually decouple 13Ca-13C\beta couplings in HNCA spectra



Hansen DF, Journal Biomolecular NMR. (2021) 75:179-191

Reconstruction NUS spectra FID-Net vs. SMILE vs. hmsIST



- 12.5% sampling
- 100 sampling schedule with a different Poisson-gap sampling schedule

Virtual Decoupling ${}^{13}C_{\alpha}$ - ${}^{13}C_{\beta}$ in 3D-HNCA and HN(CO)CA





How successful doublets are successfully decoupled yielding an improvement in resolution and two-fold increase in sensitivity for these peaks while the singlet glycine peaks are unaltered

Virtual Decoupling (II) ${}^{13}C-13C$, ${}^{13}CO-15N$, ${}^{13}C\alpha-13CO$...



Traditional methods, such as IPAP and DIPAP, require the acquisition of multiple spectra and taking linear combinations to yield a singlet resolved spectrum.

Conversely, FID-Net-based DNNs can be trained to decouple spectra with one or two couplings using a single spectrum

Virtual Decoupling (III)

¹³C-¹³C side-chain correlation spectra for per-deuterated proteins





Fig. 2 ¹³C-¹³C side-chain correlation spectra of per-deuterated proteins. **a** Schematic representation of the NMR pulse sequence used to obtain ¹³C-¹³C side-chain correlation spectra. The flow of the magnetisation between ¹³C₁ (blue) and ¹³C_p (red) is shown above the sequence with colour gradients. The following delays are used: $\Delta = 1/(4J_{CC}) \approx 7.1$ ms, $T = 1/(2J_{CC}) \approx 14.1$ ms, where J_{CC} is the one-bond ¹³C-¹³C scalar coupling constant. Rectangular pulses are high-power and not selective, bell-shaped pulses are frequency selective (90°: white outlined, 180°: black). Deuterium, ²H₁ is decoupled throughout the sequence and frequency discrimination is obtained by states-TPPI of phase $\Phi^{(3)}$. **b** Schematic representation of post-processing to obtain the decoupled spectrum. **c** Arginine ¹³C⁶-¹³C' correlation of the 18-kDa protein T4L L99A, obtained on a 1.4 mM sample at a static field of 14.1T at 278 K in 37 min



Hansen DF, Nature Communications (2019) 10:1747 Hansen DF, JACS (2021) 143:16935-42

Virtual Decoupling (IV) ¹³CO-¹⁵N correlation spectra (CON)



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FID-Net on large protein systems Methyl NMR Spectroscopy





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(a) A ¹³C-1H HSQC NMR spectra of uniformly ¹³C labelled non-deuterated proteins. (a) A ¹³C-1H HSQC NMR spectra of uniformly ¹³C labelled, non-deuterated, HDAC8 (42 kDa) processed with a standard discrete Fourier transform. (b) The spectrum in a processed with the FID-Net DNNs. (c) Comparison of FID-Net processed HSQC spectrum in b (orange) with a methyl-TROSY HMQC spectrum of an ILV specifically labelled and deuterated HDAC8 (blue). (d) A ¹³C-1H HSQC NMR spectra of uniformly ¹³C labelled MSG (80 kDa) processed with a standard discrete Fourier transform. (e) The spectrum in d processed with the FID-Net DNNs. (f) Comparison of FID-Net processed HSQC spectrum in d (orange) with a methyl-TROSY HMQC spectrum of an ILV specifically labelled and deuterated MSG (blue) for two selected regions. Many methyl groups are not visible in the ILV labelled sample, such as, Isoleucine ¹³C¹² (labelled).

FID-Net on large protein systems 3D ¹³C-HSQC-NOESY-HSQC non deuterated 80kDa MSG



312 NOE cross-peaks were observed among 170 methyl bearing residues from different regions of the protein



Figure 5: NOESY spectra of non-deuterated 80 kDa MSG. (a)-to-(**d**) 2D planes of the 3D ¹³C-¹³C-¹H NOESY spectra for methyl planes of (**a**) 15-¹³C⁵¹ and A14-¹³C⁸. (**b**) 1327-¹³C⁵¹ and A321-¹³C⁸. (**c**) A63-¹³C⁸ and L88-¹³C⁵¹. (**d**) M415-¹³C⁶ and L375-¹³C⁵¹. (**e**) Methyl groups of Ile, Leu, Val, Met, Ala, and Thr showing NOE cross-peaks in 3D ¹³C-¹³C-¹H NOESY spectra are highlighted as cyan sphere on cartoon presentation of Malate Synthase G (MSG) structure [PDB ID:1D8C]. (**f**) Normalized NOE cross-peak volumes versus interproton distances. Gray circles in the plot represent the individual data points obtained for each cross-

How to ...

- <u>https://github.com/gogulan-k/FID-Net</u>
- <u>d.hansen@ucl.ac.uk</u>

